



Title	Mycorrhizal association in <i>Pinus massoniana</i> Lamb and <i>Pinus elliottii</i> Engel inoculated with <i>Pisolithus tinctorius</i>
Author(s)	Tam, PCF
Citation	Mycorrhiza, 1994, v. 4 n. 6, p. 255-263
Issued Date	1994
URL	http://hdl.handle.net/10722/48502
Rights	Creative Commons: Attribution 3.0 Hong Kong License

Mycorrhizal associations in *Pinus massoniana* Lamb. and *Pinus elliottii* Engel. inoculated with *Pisolithus tinctorius*

Paul C. F. Tam

Department of Botany, The University of Hong Kong, Pokfulam Road, Hong Kong

Abstract. Dichotomous mycorrhizas were induced in *Pinus massoniana* and *Pinus elliottii* seedlings inoculated with *Pisolithus tinctorius* growing under non-axenic conditions. Six months after inoculation, *Pinus massoniana* seedlings exhibited a higher degree of infection, bore more mycorrhizas and had developed more abundant extramatrical mycelium than seedlings of *Pinus elliottii*. Nevertheless, seedlings of *Pinus massoniana* were stunted and exhibited chlorosis of the needles, indicating a possible nutrient deficiency. Histological examination of these pine mycorrhizas showed an ectomycorrhizal association typical of gymnosperms with an intercellular Harting net penetrating between several layers of cortical cells close to the endodermis. However, strong polyphenolic reactions, intracellular hyphae and wall modifications were occasionally observed, indicating that both host-tissue incompatibility and ectendomycorrhizal association can occur in pine species under stressed conditions.

Key words: *Pinus massoniana* – *Pinus elliottii* – *Pisolithus tinctorius* – Ectomycorrhizas – Ectendomycorrhizas

Preliminary investigations by Chan and Griffiths (1988) showed both pine species to be ectomycorrhizal formers with a number of known fungal isolates, including the broad-host fungus *Pisolithus tinctorius* (Pers.) Coker & Couch. Several pine species have been reported to develop ectomycorrhizal infections when inoculated with *Pisolithus tinctorius* in semi-axenic conditions of growth in pouches (Fortin et al. 1980; Warrington et al. 1981; Piche and Fortin 1982; Piche et al. 1982, 1983a,b), but little has been reported on ectendomycorrhizal associations under these conditions. However, the prevalence of ectendomycorrhizas in pine species infected by E-strain fungi occurring in forest nurseries has been well documented (Harley and Smith 1983). In this present investigation, the nonaxenic technique (Tam and Griffiths 1993a) was used to synthesize and develop mycorrhizas from two pine species, *Pinus massoniana* and *Pinus elliottii*, inoculated with *Pisolithus tinctorius*. The growth of host seedlings and mycorrhizal development were recorded photographically and mycorrhiza morphology and histology were examined in detail by scanning electron, and both light- and laser scan confocal microscopy.

Introduction

Pinus massoniana Lamb., 'Chinese red pine', is a dominant native conifer which has spread for centuries throughout the temperate regions of southern China and Hong Kong. *Pinus elliottii* Engel., 'slash pine', a North American conifer, was introduced into Hong Kong some five decades ago by the Agriculture and Fisheries Department during reforestation programmes. Because of their drought resistance and ability to establish on treeless hill slopes, both pine species were widely planted and are now important woodland components throughout the territory of Hong Kong.

Materials and methods

Fungal cultures

A culture of *Pisolithus tinctorius* (Pers.) Coker & Couch was obtained from the American Type Culture Collection (ATCC 38054) and maintained on modified Melin-Norkrans (MMN) agar medium.

Seedlings

Pine seeds were germinated in petri dishes containing a few pieces of moistened filter paper. Five- to 7-day-old seedlings with no visible fungal contamination were transferred to paper-wicked test tubes and grown as described previously (Tam and Griffiths 1993a). One-month old seedlings were also transferred to paper-sandwiched glass plates. These were inoculated and grown in a

115/2

compartmented thin-layer-chromatography tank under conditions described previously (Tam and Griffiths 1993a).

Growth observations

Mycorrhizal development and the growth of the seedlings in the paper-sandwiched glass plates were recorded photographically after 1-month and 6-month periods, respectively, following inoculation. Samples of short roots were removed and fixed in 4% buffered glutaraldehyde and embedded in glycol methacrylate (GMA); 2- to 3- μ m sections were cut for light and confocal microscopy. Sections for confocal microscopy were either observed unstained or were stained with 0.1% ethidium bromide in 75% alcohol for 1 min, washed with distilled water and mounted in 50% glycerol. Similar root materials were also processed for scanning electron microscopy as described previously (Tam and Griffiths 1993b).

Results

Mycorrhizal development

Seedlings of both pine species produced root systems with numerous first-order lateral roots when sandwiched between chromatography paper and glass plates (Figs. 1, 5). Seven to 14 days after inoculation, lateral roots contacting the hyphae emanating from fungal inocula were induced to form dichotomous short roots which were engulfed by bright yellow hyphal wefts. Two months later, a network of buff-brown extramatrical hyphae was formed which proliferated on the main root and bridged various mycorrhizal apices growing at different locations of the root system (Figs. 2a, b, 6a, b). After 6 months, lateral roots in contact with the fungal inocula were stimulated to form mycorrhizal clusters. Extensive networks of extramatrical hyphae, derived from the mycorrhizal apices and composed of thick mycelial strands and diffuse hyphae, colonized the root systems and interconnected with those of the neighboring seedlings (Figs. 3, 4, 7, 8).

Compared to seedlings of *Pinus elliotii*, the mycorrhizal strands of *Pinus massoniana* were more abundantly produced and the area of the root systems colonized by extramatrical hyphae was much larger; mycorrhizas synthesized in *Pinus massoniana* visibly outnumbered those formed in *Pinus elliotii* (Figs. 4, 8). At the beginning of inoculation, green needles were found on seedling shoots of both pine species. However, after 6 months, most needles, especially those of *Pinus massoniana*, appeared yellow to purple and older needles on the lower portion of the shoot became brown and withered. With regard to the overall growth pattern of seedlings of the two pine species, *Pinus elliotii* exhibited a significant increase in shoot height after 6 months, whereas *Pinus massoniana* showed no shoot height increment even after 5 months. However, seedlings of both pine species after 6 months produced extensive root growth on the chromatography paper.

Scanning electron microscopy

In the early stages of mycorrhiza formation, mantles were formed on the first-order lateral roots behind the meristemic apex (Fig. 9). Emerging hyphae, derived from the inner mantle enclosing the base of the lateral root, colonized the isodiametric apical cells and a few highly branched hyphae were seen tightly appressed to the apical surface (Fig. 10). The inner mantles of the subapical regions were made up of interwoven hyphae embedded in an amorphous mucilaginous substance (Fig. 11). With increased hyphal ramification on the apical surface, a new mantle composed of interwoven hyphae enclosing the apex and strands of five to seven hyphae grew from the loosely bound outer mantle (Fig. 12).

Light microscopy

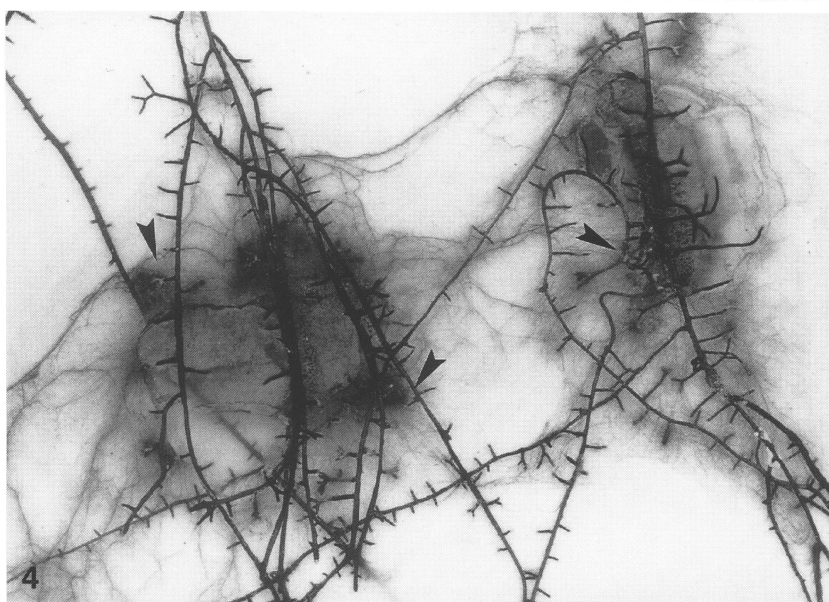
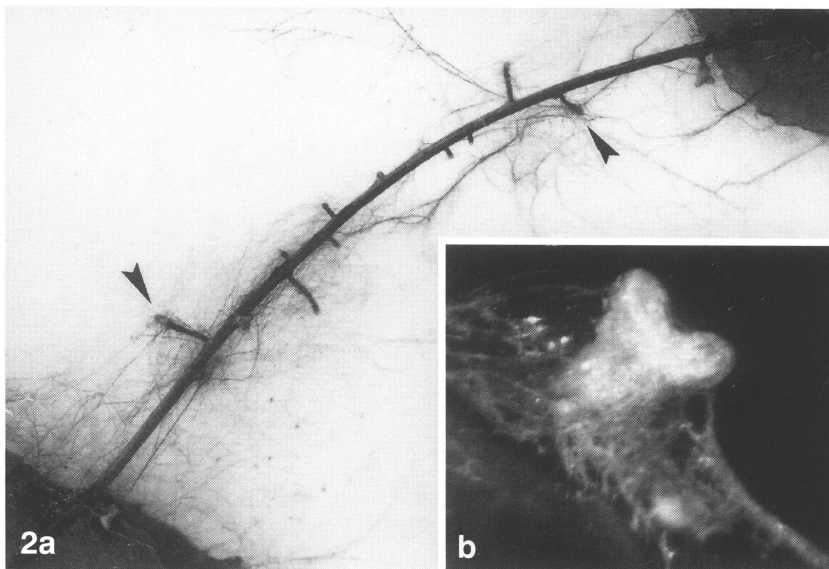
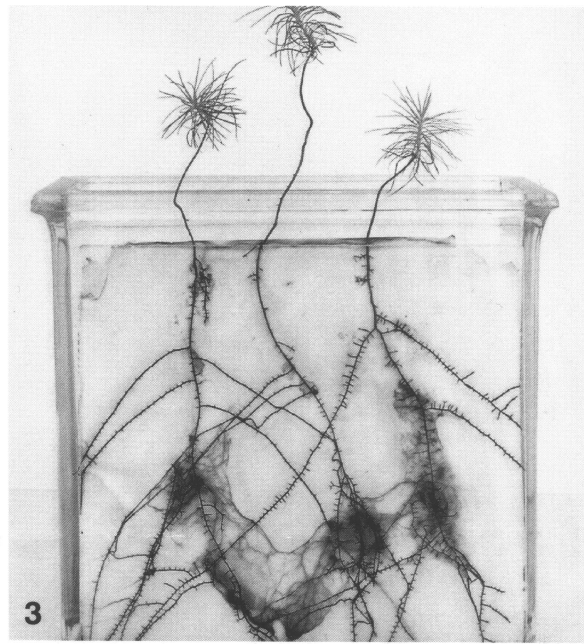
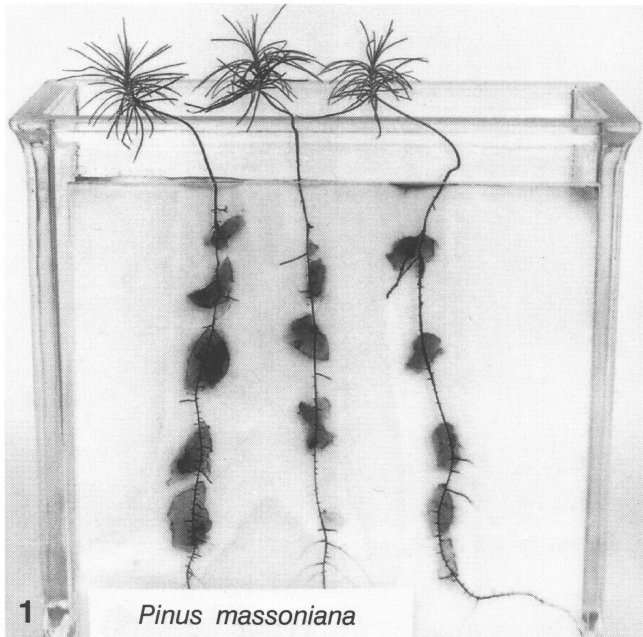
GMA longitudinal sections of synthesized mycorrhizas of both pine species with *Pisolithus tinctorius* stained with toluidine blue showed the histological features of two different types of mycorrhizal association.

Ectomycorrhizal association. Ectomycorrhizas were composed of a uniform mantle, 20–30 μ m thick, with an outer layer of prosenchymatous hyphae and an inner layer of synenchymatous hyphae; the intercellular hyphae of the Harting net penetrated three to four layers of cortical cells close to the endodermis. The nuclei appeared large and somewhat convoluted at the nuclear periphery (Figs. 13, 14).

Ectendomycorrhizal association. Ectendomycorrhizas were composed of an irregularly thick mantle, 20–50 μ m, thicker towards the base of the lateral root. Strong intracellular polyphenolic reactions occurred in the first and second layers of cortical cells, which were in close contact with the inner mantle hyphae, and a limited Hartig net development was observed in some of this region (Fig. 15). In the area of a Hartig net, intracellular hyphae were often observed and the nuclei were either absent or found to be disintegrated inside the cortical cells. Vesicle-like bodies were frequently associated with the inner cortical walls, which were thickly impregnated with polyphenols (Fig. 16).

Laser scan confocal microscopy

Unstained longitudinal GMA sections showing the outer loosely bound mantle hyphae, closely packed inner mantle hyphae and the Harting net were autofluorescent, while the dark nuclei were conspicuous against the fluorescent polyphenolic background (Fig. 17). The vesicle-like bodies associated with the inner cortical cell wall were strongly autofluorescent, whereas the disintegrated nucleus and the intracellular hyphae were not (Figs. 18, 19). In contrast, sections stained with ethidium bromide showed fluorescent, fan-like



Figs. 1–4. External morphology of mycorrhizal associations of *Pinus massoniana* seedlings infected with *Pisolithus tinctorius*

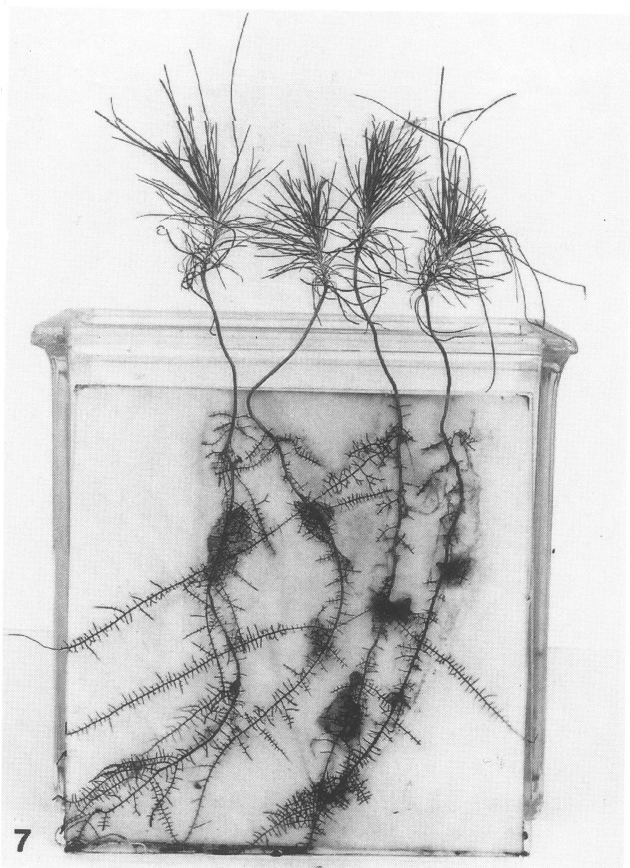
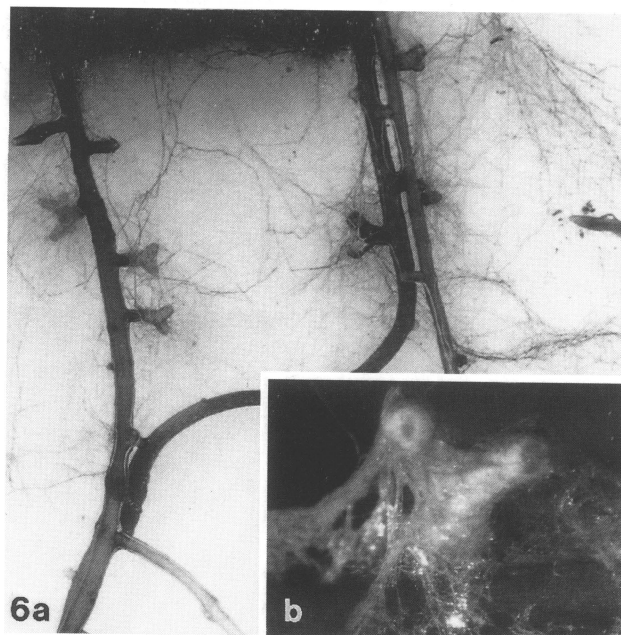
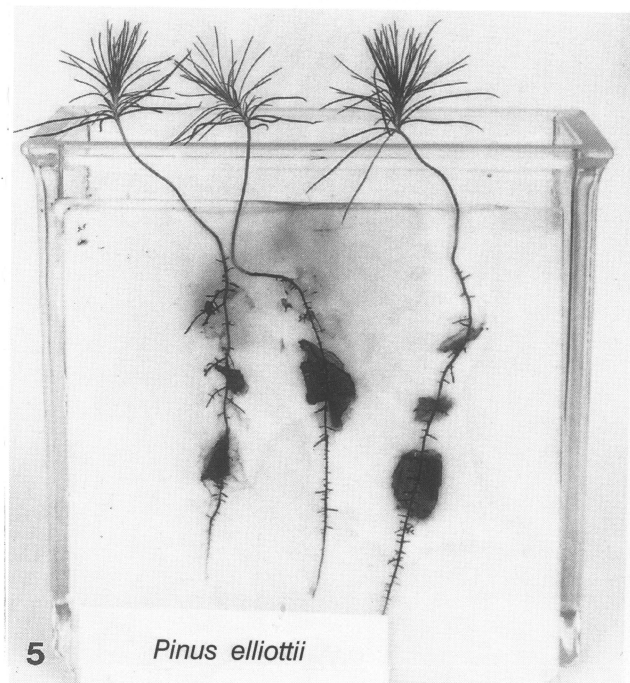
Fig. 1. One-month-old inoculated *Pinus massoniana* seedlings sandwiched between a few pieces of chromatography paper and a glass plate. $\times 1/3$

Fig. 2. a A network of extramatrical hyphal strands link various mycorrhizal apices (arrows). $\times 5$. **b** Detailed structure of dichotomous mycorrhizal roots enshrouded by hyphal weft. $\times 40$

Fig. 3. Mycelial fans developed from mycorrhizal laterals proliferate and interconnect the neighboring root systems. $\times 1/3$

Fig. 4. Detailed structure of the mycelial fan composed of a network of thicker hyphal strands and diffuse hyphae bridging various mycorrhizal apices (arrows). $\times 2$

115/4



Figs. 5-8. External morphology of mycorrhizal associations of *Pinus elliotii* seedlings infected with *Pisolithus tinctorius*

Fig. 5. One-month-old inoculated *Pinus elliotii* seedlings sandwiched between a few pieces of chromatography paper and a glass plate. $\times 1/3$

Fig. 6. a A network of extramatrical hyphal strands linking various mycorrhizal apices. **b** Detailed structure of dichotomous mycorrhizal roots enshrouded by hyphal web. $\times 40$

Fig. 7. Proliferation of mycelial strands linking neighboring root systems. $\times 1/3$

Fig. 8. Detailed structure of the mycelial strands and mycorrhizal cluster (arrow) contacting the fungal inoculum

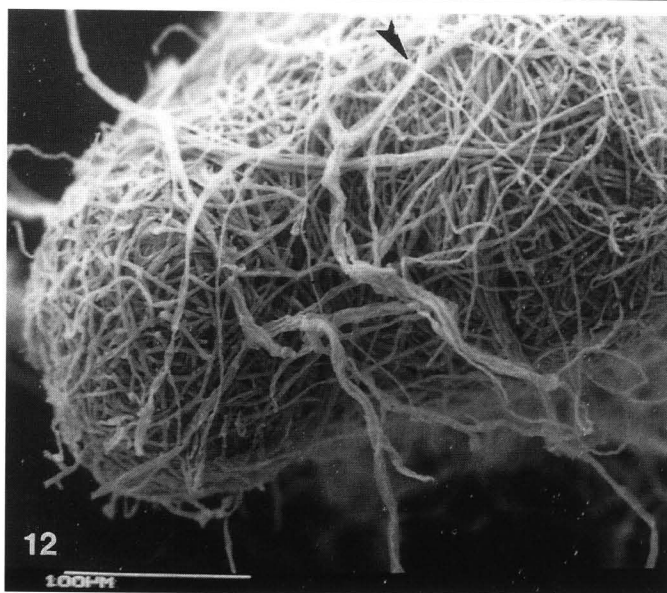
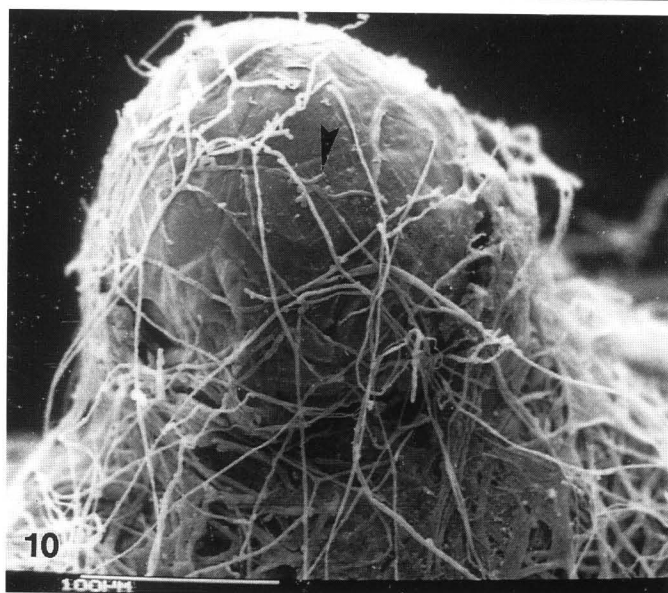
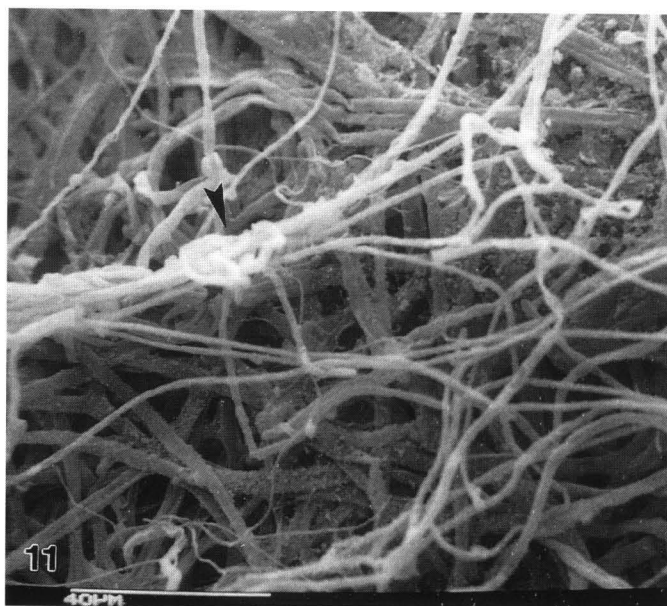
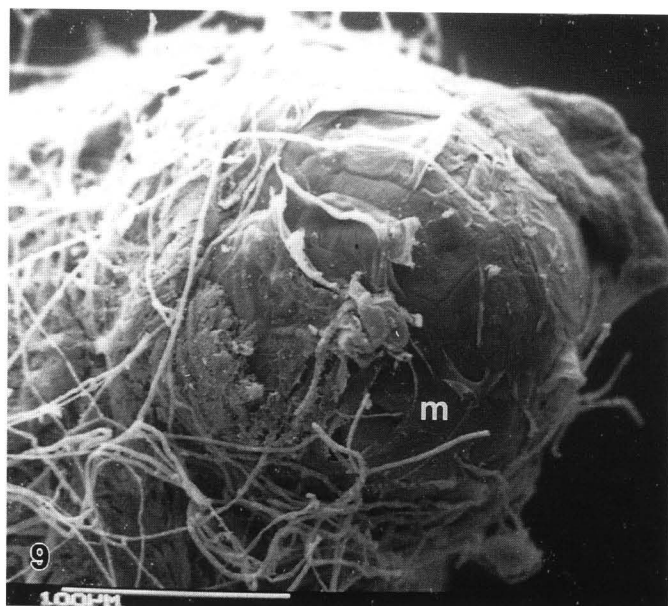


Fig. 9-12. Scanning electron micrographs showing sequence of sheath formation on the apical region of *Pinus massoniana* by *Pisolithus tinctorius*

Fig. 9. Mantle formed on the lateral root behind the apex consisting of isodiametric cells covered by a layer of mucilage-like material (m)

Fig. 10. Hyphae with clamp connections constituting a loose net on the apical surface and some highly branched hyphae (arrow) firmly cemented to the apical cells

Fig. 11. Enlarged portion of the mantle showing the interwoven hyphae embedded in mucilage-like material and loosely connected hyphal strands, branched and ornamented (arrow)

Fig. 12. The root apex completely enshrouded with new mantle and extramatrical hyphal strands (arrow) emanating from the mantle

nuclei inside the cell enclosed by the illuminated intercellular Hartig net; the cortical walls were unstained and appeared dark (Fig. 20). Stained sections also showed that intercellular hyphae had broken through the thinner modified wall and entered into the highly vacuolated cell lumen, where the brightly illuminated intracellular hyphae were also visible (Fig. 21). Wall modifications included wall thickenings, disruption, as-

sociated bodies of various shapes and vesicle-like bodies (Fig. 22).

Discussion

Seedlings of *Pinus massoniana* and *Pinus elliottii* were grown under identical conditions. Dichotomous mycor-

rhizas, which are a characteristic feature of ectomycorrhizal associations of conifer species, were induced. These results indicate the host compatibility of these two pine species to the mycobiont *Pisolithus tinctorius*. However, 6 months after inoculation, the growth and mycorrhizal development in seedlings of the two species varied. More mycorrhizas were induced and a greater quantity of extramatrical mycelium proliferated in seedlings of *Pinus massoniana* than *Pinus elliotii*, implying that the degree of host compatibility of the same fungus may vary with the plant species, even within the same genus. After 6 months, *Pinus massoniana* seedlings were stunted and many needles had withered and dropped, indicating a nutrient deficiency. This can be attributed to enhanced mycorrhizal infection of the root system and diversion of photosynthate from the host to the mycosymbiont for mycelial growth. Reductions in plant dry weight due to mycorrhizal infections have been reported previously (Reid et al. 1983; Cairney et al. 1989; Dosskey et al. 1990), and a negative correlation between the degree of fungal development in terms of extramatrical mycelium and the growth of the host plants, as demonstrated in the present study, was also shown by Colpaert et al. (1992). Extramatrical mycelial strands serve as channels for the translocation of water and mineral nutrients (Read and Boyd 1986), and mycelial networks linked to root systems of the same or another plant species may have physiological and ecological roles (Read 1976; Allen 1991).

The formation of dichotomous mycorrhizas in these two pine species is similar to those described previously (Piche et al. 1982, 1983a), and amorphous mucilage-like material secreted by the host root may be responsible for an initial step in ectomycorrhizal attachment or may act as a recognition factor (Piche et al. 1983a).

Duddridge and Read (1984a,b) suggested that one of the most striking features of the host-fungus interface in ectomycorrhizal associations is the apparent absence of cytoplasmic reaction to the presence of intercellular fungus. In the present study, mycorrhizal material also showed a regular intercellular Hartig net enclosing the host cytoplasm with large nuclei but without an aggressive intracellular polyphenolic response, indicating some mutualistic feature of ectomycorrhizal association. Marks and Foster (1973) and Marx (1972) suggested that the presence of polyphenols or the availability of soluble sugars in the cortical cells contacting mantle and Hartig net hyphae in ectomycorrhizas inhibits fungal cellulase production, thus restricting intracellular penetration. However, in the present study, strong intracellular polyphenolic reactions were also observed at the host-fungus interface even with limited Hartig net development. This reaction indicates a hypersensitive response by the host towards the incompatible mycorrhizal fungus (Malajczuk et al. 1984; Giovannetti and Lioi 1990) and is analogous to the host response towards pathogens. The ectomycorrhizal fungus was demonstrated to have the ability to penetrate highly modified walls into the senescing cor-

tical cells and produce ectendomycorrhizas. This ectendomycorrhizal condition may have been induced by the limited carbohydrate supply from the host arising from stunted shoot growth and defoliation, itself a result of the high degree of mycorrhizal colonization by extramatrical hyphae. The production of cellulase is positively correlated with the glucose content of the root (Melin 1948, 1953), and lytic enzymes produced by the mycosymbiont may lead to wall destruction and intracellular hyphal formation as well as wall modifications such as vesicle-like bodies. Similar wall bodies have been observed in mycorrhizal roots of *Pinus mugo* (Wills and Cole 1978), and modified wall structures such as papillae and wall ingrowth occurred in *Picea abies* (Nylund et al. 1982; Kottke and Oberwinkler 1986), while wall protuberances were observed in natural pine species (Duddridge and Read 1984a). All such wall modifications are probably brought about by host-wall/fungus interactions.

Wilcox (1983) suggested that the ectomycorrhizal condition is not absolute and can be influenced towards the ectendomycorrhizal condition by either associant; similarly, ectomycorrhizal fungi can also change from beneficial to harmful under some conditions of stress. The shift of pine mycorrhizas infected with *Pisolithus tinctorius* from a mutualistic, beneficial ectomycorrhizal association to a harmful, saprophytic- or parasitic-like ectendomycorrhizal association, probably the result of the plant host losing control of the mycobiont, is clearly evident in this present study.

Acknowledgements. The author wishes to express his thanks to Professor D. A. Griffiths for critically reading the manuscript and Mr. Jason W. T. Tam for his help with the confocal microscopy.

Figs. 13–16. Light micrographs of longitudinal glycol methacrylate (GMA) sections of synthesized mycorrhizal root *Pinus elliotii*/*Pisolithus tinctorius*

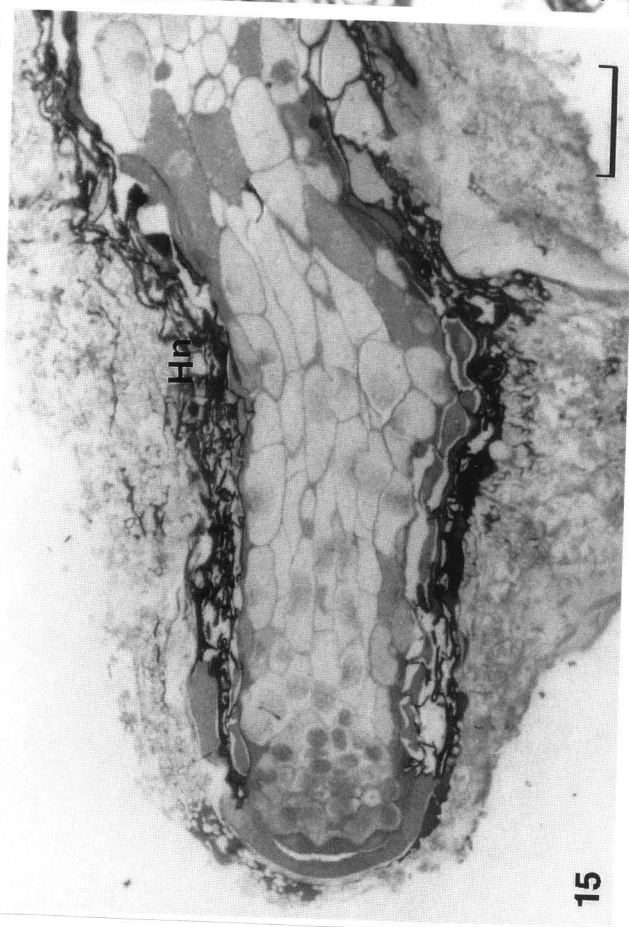
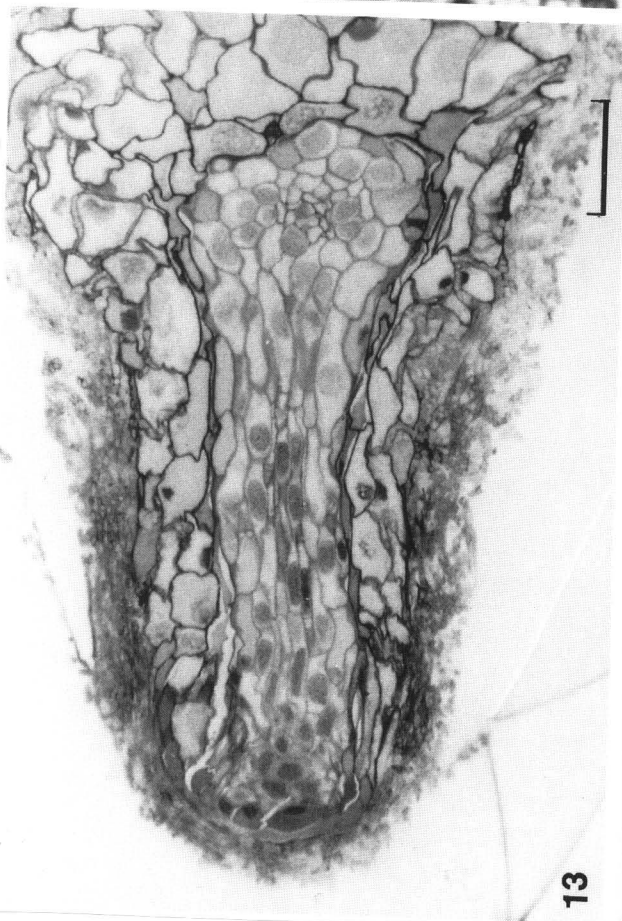
Fig. 13. Mycorrhizal root composed of outer prosenchyma and inner synenchyma and intercellular Hartig net extending through several layers of cortical cells close to the endodermis; bar = 50 µm

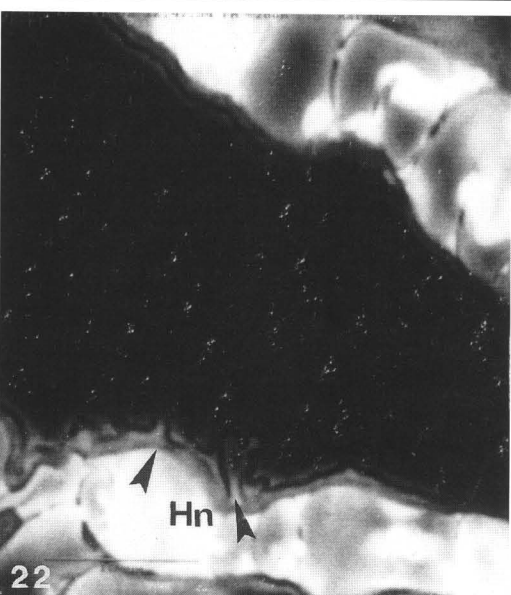
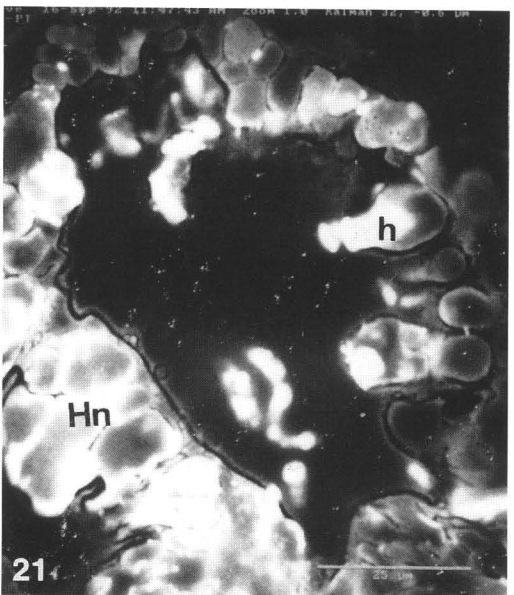
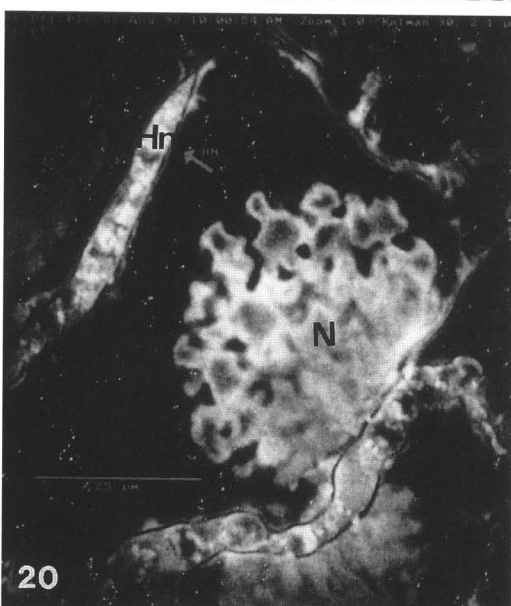
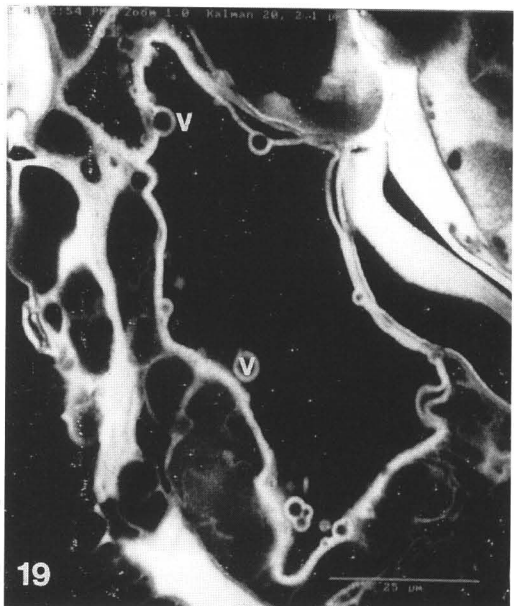
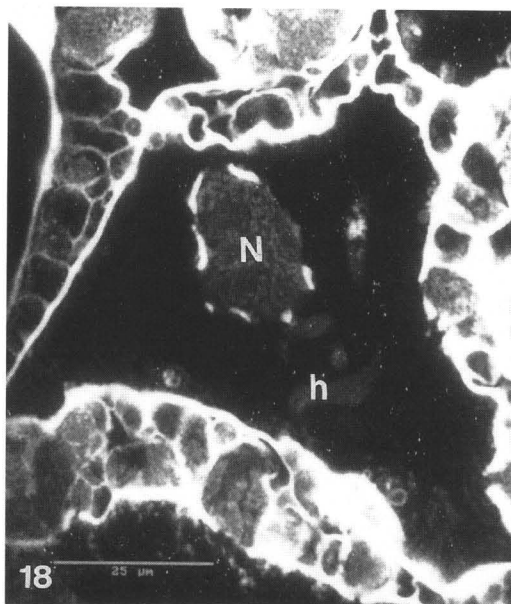
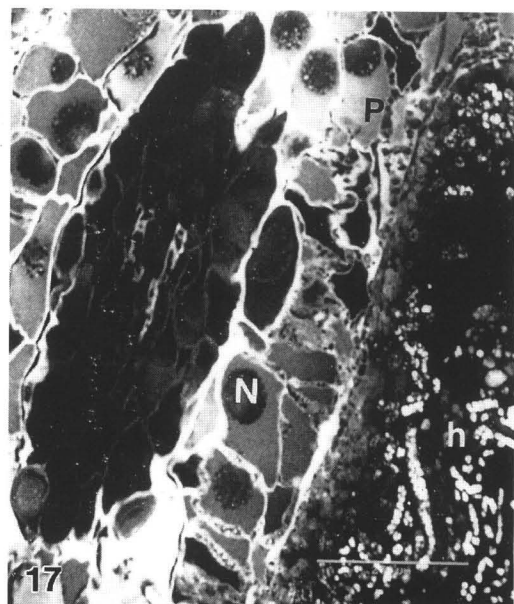
Fig. 14. Enlarged portion of the intercellular Hartig net and conspicuous, convoluted nuclei (N) inside cortical cells; bar = 25 µm

Fig. 15. Mycorrhizal root with thick mantle. Note strong, dark polyphenolic reaction (arrow) and limited Hartig net (Hn) development; bar = 50 µm

Fig. 16. Enlarged portion of the cortical cells showing intercellular Hartig net (Hn), disintegrated nucleus (N) and intracellular hyphae (h). Note polyphenol-impregnated cell walls (arrow) and vesicle-like bodies (v); bar = 25 µm

115/7





Figs. 17–22. Laser scan confocal micrographs of GMA sections of synthesized mycorrhizal root *Pinus elliotii*/*Pisolithus tinctorius*. **Figs. 17–19** are unstained sections and **Figs. 20–22** are stained sections

Fig. 17. Mycorrhiza showing fluorescent outer mantle hyphae (*h*), polyphenol-containing cells (*P*), cortical walls and Hartig net (*Hn*) and dark nuclei (*N*)

Fig. 18. Detailed structure of the mycorrhiza showing autofluorescent cortical walls in contrast to dimly illuminated intracellular hyphae (*h*) and the deformed nucleus (*N*)

Fig. 19. Detailed structure of cortical cells showing fluorescent, vesicle-like bodies (*v*) attached to the walls

Fig. 20. Detailed structure of the mycorrhiza showing fluorescent intercellular Hartig net (*Hn*) hyphae confined to dark cortical walls and a fluorescent fan-shaped nucleus (*N*)

Fig. 21. Detailed structure of the mycorrhiza showing fluorescent intercellular hyphae (*h*) breaking through disrupted cortical walls and entering into the cell lumen as intracellular hyphae (*h*)

Fig. 22. Detailed structure of wall modifications showing various-shaped bodies (*arrows*) associated with the cortical walls

References

- Allen MF (1991) The ecology of mycorrhizae. Cambridge University Press, Cambridge, UK
- Cairney WG, Ashford AE, Allaway WG (1989) Distribution of photosynthetically fixed carbon within root systems of *Eucalyptus pilularis* plants ectomycorrhizal with *Pisolithus tinctorius*. New Phytol 112:495-500
- Chan WK, Griffiths DA (1988) The mycorrhizae of *Pinus elliottii* Engel. and *P. massoniana* Lamb. in Hong Kong. Mem Hong Kong Nat Hist Soc 18:11-17
- Colpaert JV, Assche JAV, Luijters K (1992) The growth of extramatrical mycelium of ectomycorrhizal fungi and the growth response of *Pinus sylvestris* L. New Phytol 120:127-135
- Dosskey MG, Linderman RG, Boersma L (1990) Carbon-sink stimulation of photosynthesis in Douglas fir seedlings by some ectomycorrhizas. New Phytol 115:269-274
- Duddridge JA, Read DJ (1984a) The development and ultrastructure of ectomycorrhizas. I. Ectomycorrhizal development on pine in the field. New Phytol 96:565-573
- Duddridge JA, Read DJ (1984b) The development and ultrastructure of ectomycorrhizas. II. Ectomycorrhizal development of pine in vitro. New Phytol 96:575-578
- Fortin JA, Piche Y, Lalonde M (1980) Technique for observation of early morphological changes during ectomycorrhiza formation. Can J Bot 58:361-365
- Giovannetti M, Lioi L (1990) The mycorrhizal status of *Arbutus unedo* in relation to compatible and incompatible fungi. Can J Bot 68:1239-1244
- Kottke I, Oberwinkler F (1986) Root-fungus interactions observed on initial stages of mantle formation and Hartig net establishment in mycorrhizas of *Amanita muscaria* on *Picea abies* in pure culture. Can J Bot 64:2348-2354
- Malajczuk N, Molina R, Trappe JM (1984) Ectomycorrhiza formation in *Eucalyptus*. II. The ultrastructure of compatible and incompatible mycorrhizal fungi and associated roots. New Phytol 96:43-53
- Marks GS, Foster RC (1973) Structure, morphogenesis, and ultrastructure of ectomycorrhizae. In: Marks GC, Kozlowski TT (eds) Ectomycorrhizae, their ecology and physiology. Academic Press, New York, pp 1-41
- Marx DH (1972) Ectomycorrhizae as biological deterrents to pathogenic root infections. Annu Rev Phytopathol 10:429-454
- Melin E (1948) Recent advances in the study of tree mycorrhiza. Trans Br Mycol Soc 30:92-99
- Melin E (1953) Physiology of mycorrhizal relations in plants. Annu Rev Plant Physiol 4:325-346
- Nylund JR, Kasimir A, Arveby SS (1982) Cell wall penetration and papilla formation in senescent cortical cells during ectomycorrhizal synthesis in vitro. Physiol Plant Pathol 21:71-73
- Piche Y, Fortin JA (1982) Development of mycorrhizae, extramatrical mycelium and sclerotia on *Pinus strobus* seedlings. New Phytol 91:211-220
- Piche Y, Fortin JA, Peterson RL, Posluszny U (1982) Ontogeny of dictotomizing apices in mycorrhizal short roots of *Pinus strobus*. Can J Bot 60:1523-1528
- Piche Y, Peterson RL, Ackerley CA (1983a) Early development of ectomycorrhizal short roots of pine. Scanning Electron Microsc 3:1467-1474
- Piche Y, Peterson RL, Howarth MJ, Fortin JA (1983b) A structural study of the interaction between the ectomycorrhizal fungus *Pisolithus tinctorius* and *Pinus strobus* roots. Can J Bot 61:1185-1193
- Read DJ (1976) The structure and function of vegetative mycelium of mycorrhizal roots. In: Jennings DH, Rayner ADM (eds) The ecology and physiology of fungal mycelium. Cambridge University Press, Cambridge, UK, pp 215-240
- Read DJ, Boyd R (1986) Water relations of mycorrhizal fungi and their host plants. In: Ayres PG, Boddy L (eds) Water, fungi and plants. Cambridge University Press, Cambridge, UK, pp 287-303
- Reid CPP, Kidd FA, Ekwebelam SA (1983) Nitrogen nutrition, Photosynthesis and carbon allocation in ectomycorrhizal pine. Plant Soil 71:415-432
- Tam PCF, Griffiths DA (1993a) Mycorrhizal associations in Hong Kong Fagaceae. I. Techniques for rapid detection and observation of ectomycorrhizas in local genera. Mycorrhiza 2:111-115
- Tam PCF, Griffiths DA (1993b) Mycorrhizal associations in Hong Kong Fagaceae. III. The ontogeny of mycorrhizal development, growth and nutrient uptake by *Quercus myrsinaefolia* seedlings inoculated with *Pisolithus tinctorius*. Mycorrhiza 2:125-131
- Warrington SJ, Black HD, Coons LB (1981) Entry of *Pisolithus tinctorius* hyphae into *Pinus taeda* roots. Can J Bot 59:2135-2139
- Wilcox HE (1983) Fungal parasitism of woody plant roots from mycorrhizal relationship to plant disease. Annu Rev Phytopathol 21:221-242
- Wills BJ, Cole LJ (1978) A scanning microscopy study of 'vesicular bodies' in mycorrhizal roots of *Pinus mugo* (Turra). New Phytol 80:579-582

Harley JL, Smith SE (1983) Mycorrhizal symbiosis
Academic Press, London, New York